

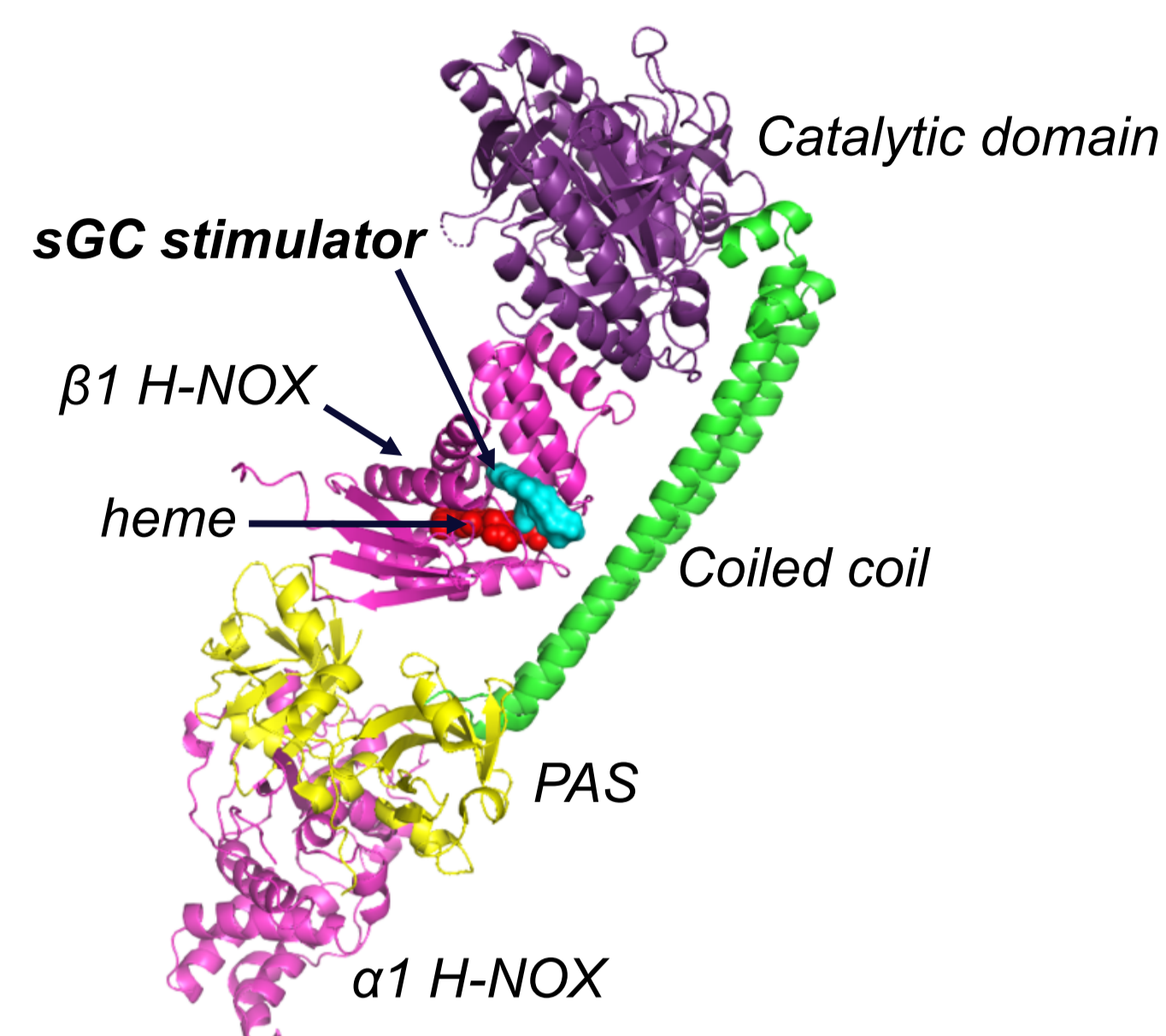
Development of a Soluble Guanylate Cyclase Radioligand Binding Assay Using [³H]-Praliguat

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Introduction

Soluble guanylate cyclase (sGC) stimulators are a class of small molecule agonists that bind to sGC and act in synergy with nitric oxide (NO) to increase production of cGMP from GTP. sGC stimulators have been characterized for their ability to stimulate cGMP production in cellular and purified enzyme assays, but a robust assay for determining relative binding affinities of sGC stimulators has been lacking. Praliguat is an investigational, oral, once-daily sGC stimulator.

Structural model of stimulator bound to sGC



Presented by Jung et al.,
8th International Conference on cGMP (2017)

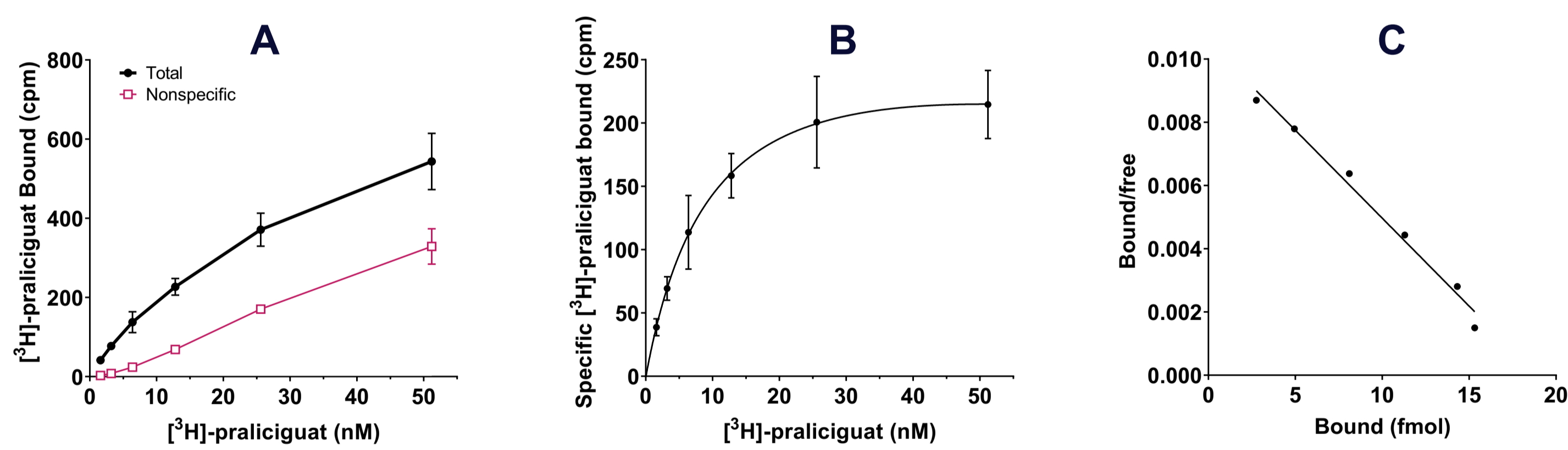
Methods

We adapted a binding assay using [³H]-praliguat radioligand and size-exclusion chromatography to analyze the binding of praliguat to purified human recombinant sGC. [³H]-praliguat was custom synthesized by American Radiolabeled Chemicals (St. Louis, MO) with a specific activity of 14 Ci/mmol. NO was provided as DETA-NO. The binding assay was used to explore cofactors required for binding and was further developed as a competitive binding assay.

Results

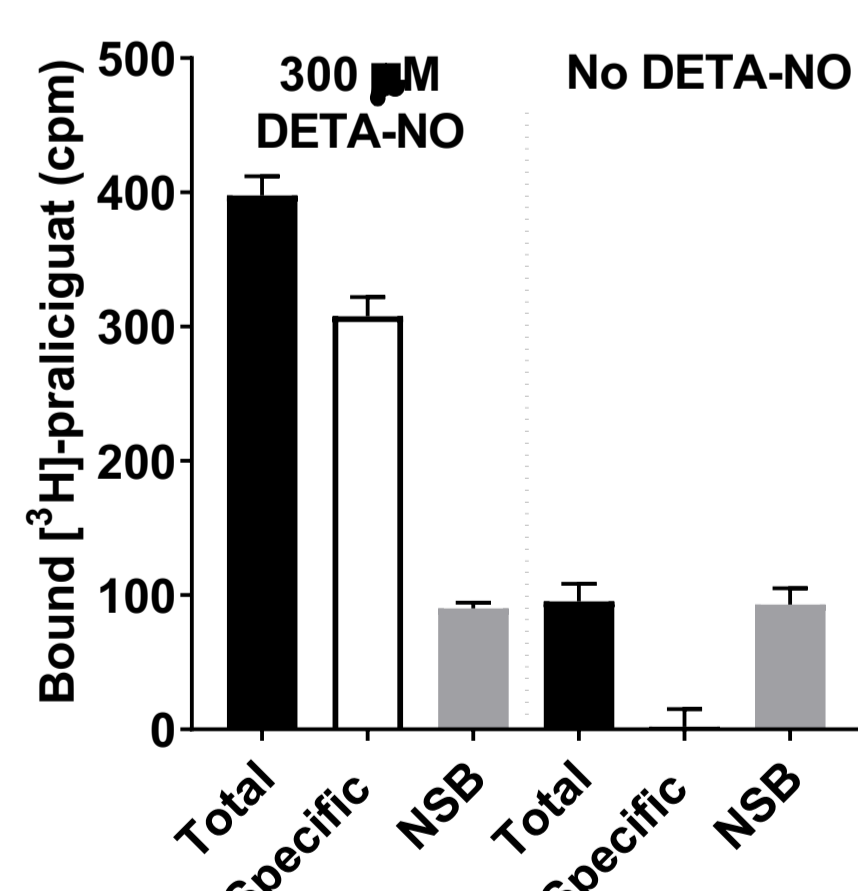
[³H]-Praliguat bound to human recombinant sGC with high affinity

- Saturable binding with 6-9 nM K_D
- No cooperativity, single binding site (linear Scatchard)



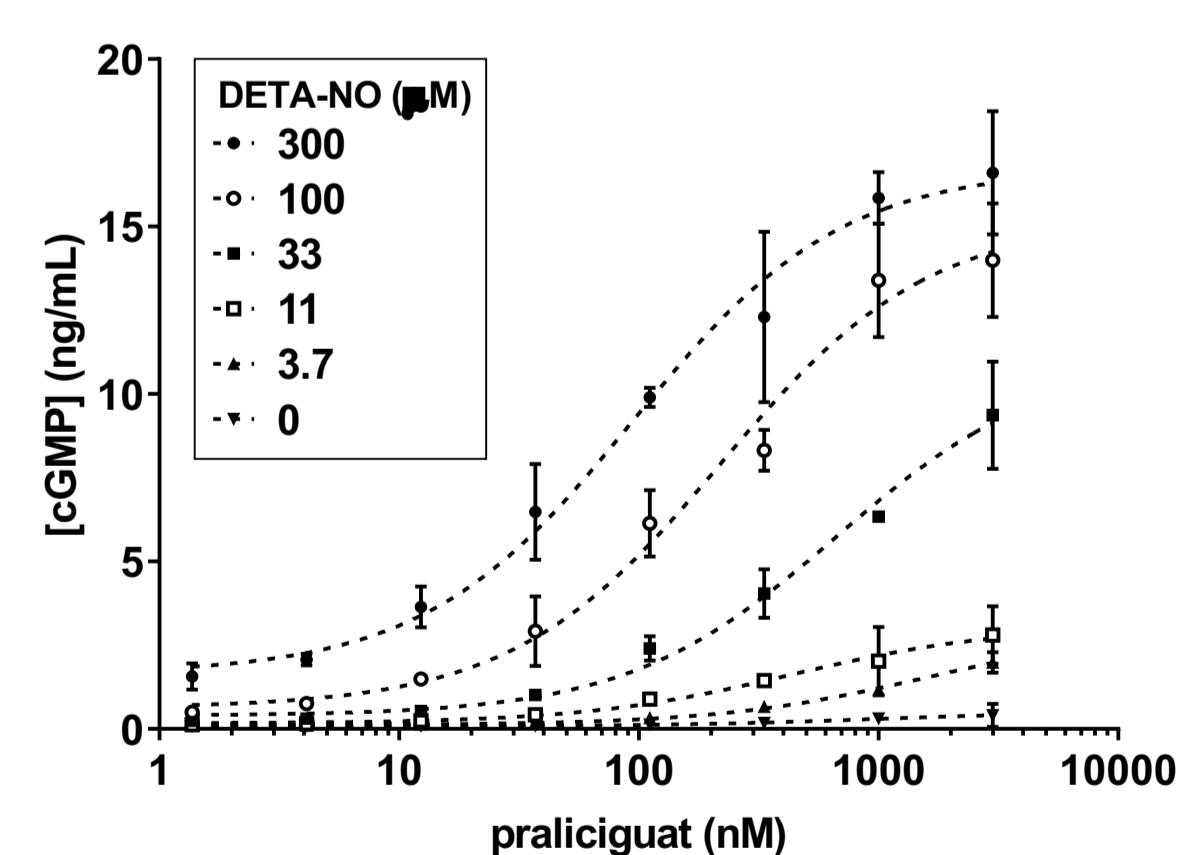
Binding conditions: 40 ng sGC (1.3 nM), 0.3 mM DETA-NO, 0.4 mM GTP, 1.6-51.2 nM [³H]-praliguat (n=3 replicates). **A.** [³H]-praliguat, in the presence (Nonspecific) and absence (Total) of 1 μM praliguat for 22 h at 37°C. **B.** Specific saturation binding curve yield a 6.9 nM K_D and a Hill slope of 1.1. **C.** Scatchard plot resulted in a 9 nM K_D . Under conditions similar to those used for binding, sGC was fully active with a cGMP-forming activity of 10.8 μmol cGMP/min/mg protein (assay conditions: 40 ng sGC, 0.3 mM DETA-NO, 3 mM MgCl₂, 0.4 mM GTP, 2 mM DTT, 0.025% BSA; 30 min at room temperature).

Praliguat binding required NO



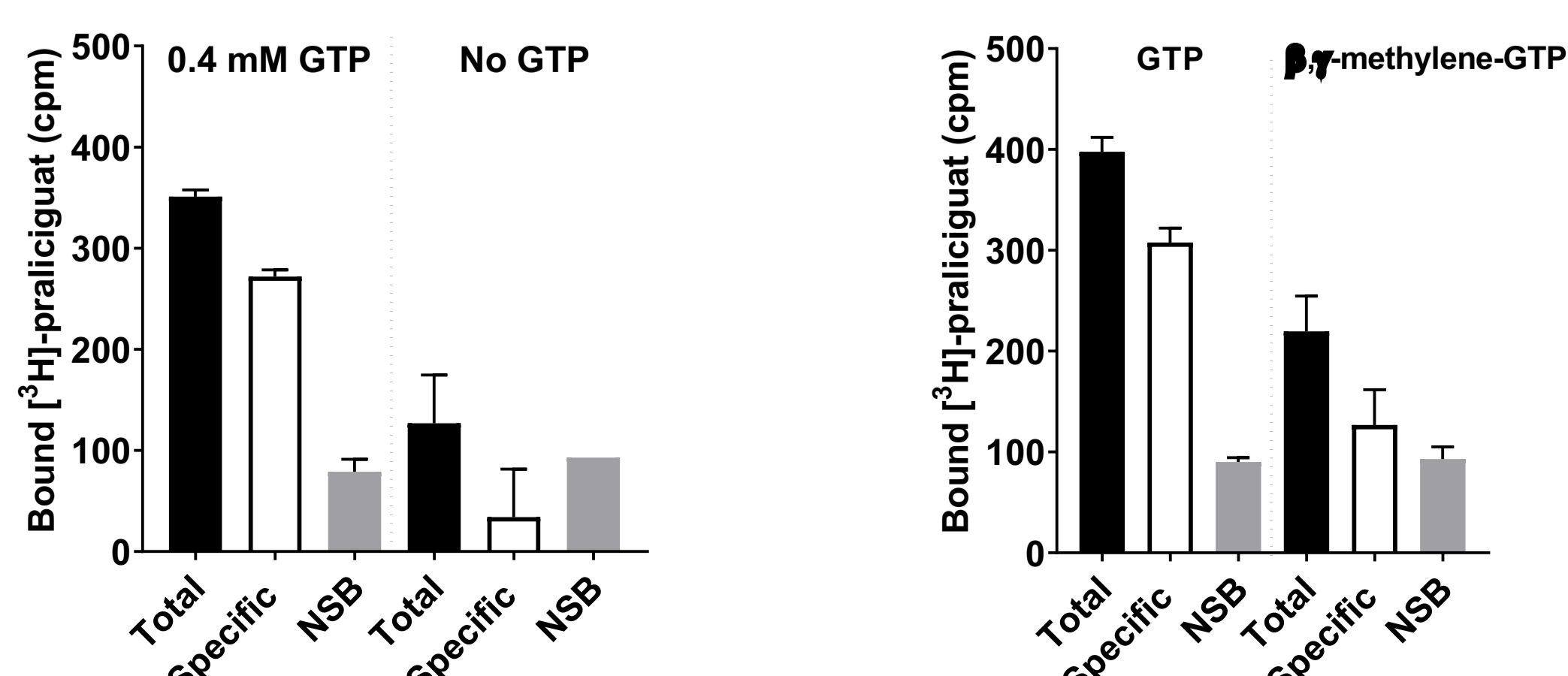
Binding of [³H]-praliguat to sGC in the presence and absence of DETA-NO

Praliguat was synergistic with NO in the sGC activity assay



Praliguat concentration response in human recombinant sGC activity assay under increasing concentrations of DETA-NO (0 μM to 300 μM).

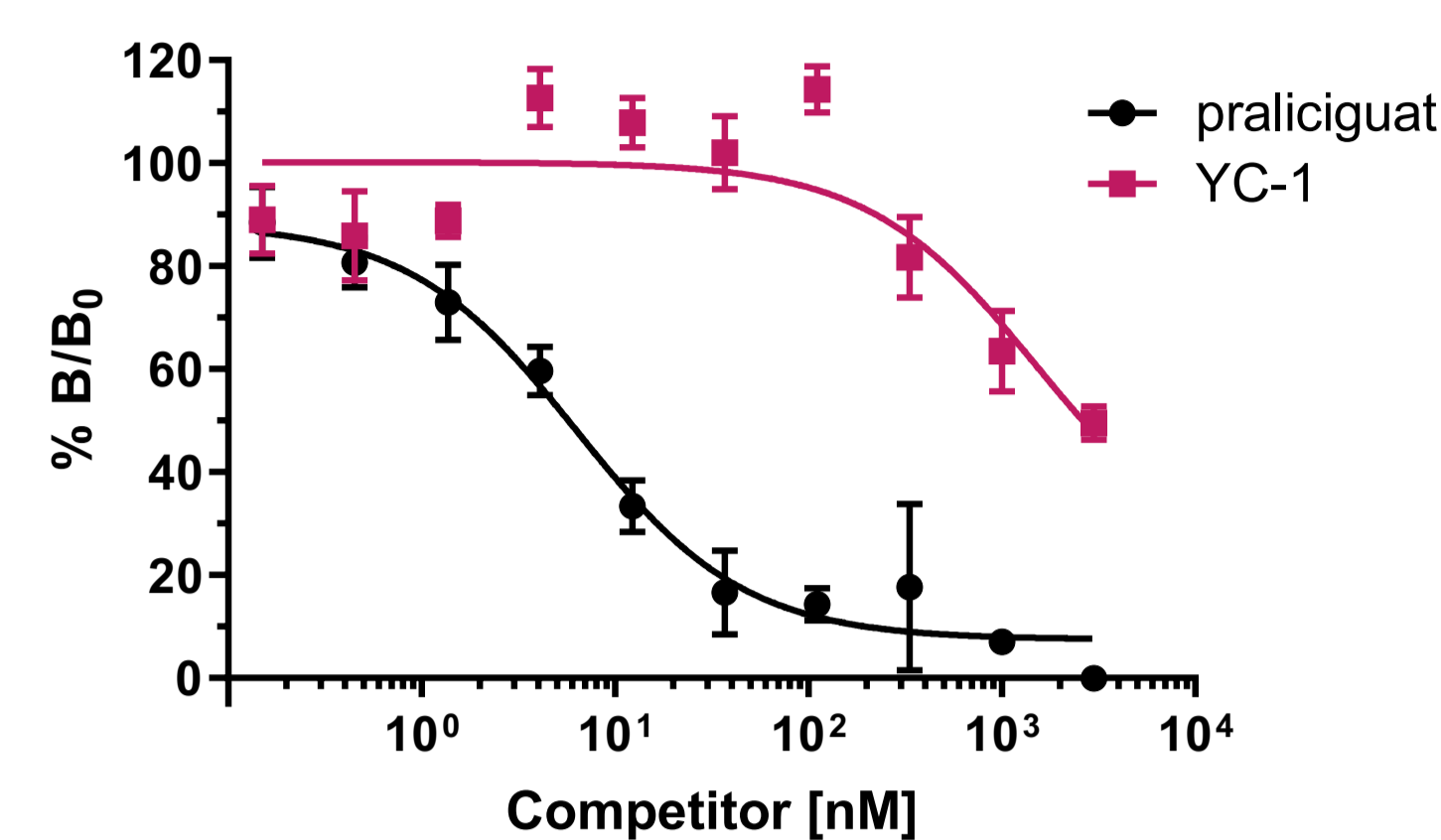
Praliguat binding was greater with GTP



Binding of [³H]-praliguat to sGC in the presence and absence of GTP and nonhydrolyzable GTP analogs (0.4 mM).

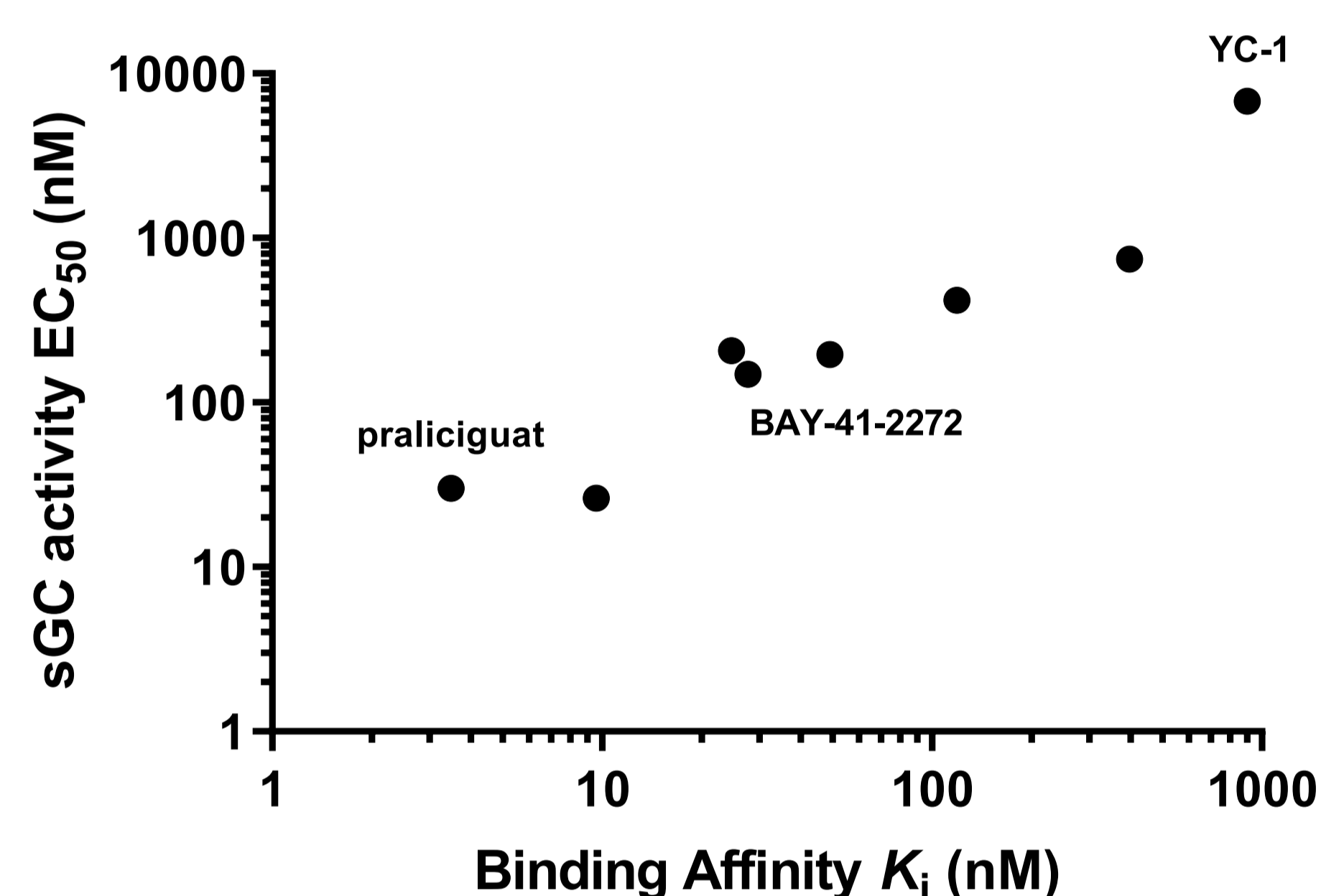
Results (continued)

Competitive radioligand binding



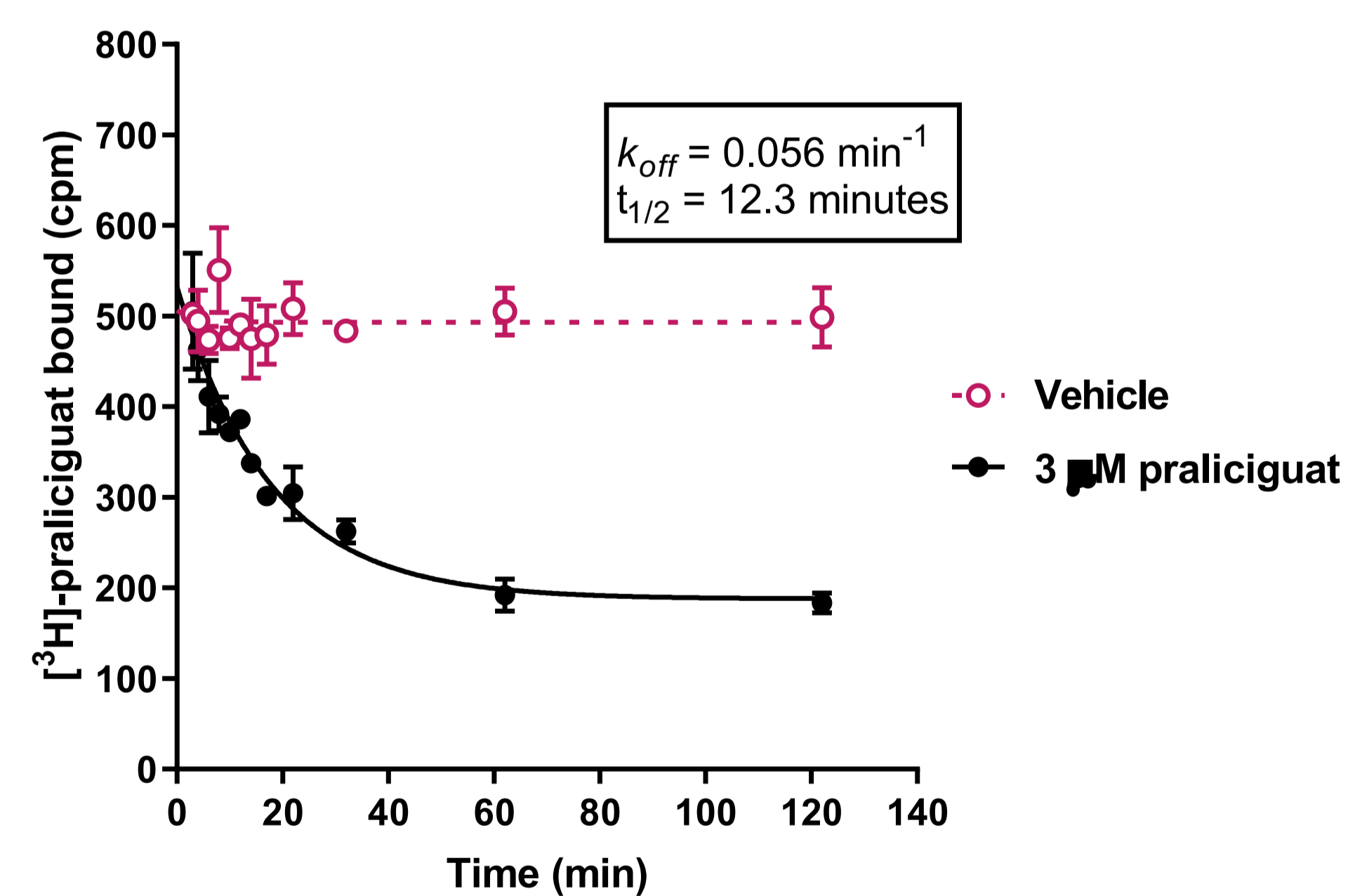
Competitive radioligand-binding curve using [³H]-praliguat. Human recombinant sGC was incubated with [³H]-praliguat and increasing concentrations of unlabeled praliguat or the sGC stimulator YC-1 for 1.5 h at 37°C. sGC was isolated using gel filtration chromatography and radioactivity was quantitated in a liquid scintillation counter. The K_i for praliguat was 3.64 ± 0.26 nM (n=4 experiments). %B/B₀ was calculated as bound radioactivity in the presence of competitor relative to bound radioactivity determined with no competitor.

Binding affinity correlated with potency in cGMP assay



Correlation between relative binding affinities (K_i) of different sGC stimulators in the competitive radioligand binding assay and potency (EC_{50}) in the human sGC activity assay performed in the presence of 30 μM DETA-NO.

In a kinetic assay, the dissociation half-life of praliguat from sGC was 12.3 minutes



Kinetics of [³H]-praliguat dissociation from sGC. Human recombinant sGC was incubated with [³H]-praliguat for 1 h at 37°C followed by challenge with either vehicle (DMSO) or 3 μM praliguat. [³H]-Praliguat bound to sGC was isolated at different times after the challenge using gel filtration chromatography and quantitated in a liquid scintillation counter. The calculated association rate (assuming $K_D=9$ nM and $k_{off}=0.056$ min⁻¹) was 6.2 X 10⁶ M⁻¹ min⁻¹.

Conclusions

- Praliguat bound with high affinity to a single binding site on sGC with no observed cooperativity.
- In the binding assay, praliguat binding required NO and was greater in the presence of the sGC substrate, GTP.
- Our observations are consistent with the role of praliguat as an allosteric modulator of sGC, potentiating the activity of NO.
- This binding assay can be used to explore binding kinetics and may be a useful tool for identification and characterization of sGC stimulators with novel binding and kinetic properties.